

# **PathoSEEK<sup>®</sup> Pseudomonas aeruginosa and Staphylococcus aureus Multiplex Detection Assay v2 with MaGiC Lysis Kit**

## **Method Developer Validation**

**Real Time PCR (qPCR) assay for the detection of *Pseudomonas aeruginosa* and  
*Staphylococcus aureus* in dried cannabis flower and infused products matrices**

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## **Abstract**

### ***Background:***

Current regulations require cannabis flower and cannabis products to be free of species of *Pseudomonas aeruginosa* and *Staphylococcus aureus*. The PathoSEEK® *Pseudomonas aeruginosa* and *Staphylococcus aureus* Multiplex Detection Assay v2 with MaGiC Lysis Kit will detect these species on two separate optical channels using a qPCR (Quantitative Polymerase Chain Reaction) assay allowing for speciation of the two in cannabis flower, cannabis concentrates, infused edibles and infused non-edibles. An Optional Grim Reeper (free DNA removal) step may be employed to reduce positive results caused by non-viable target organisms.

### ***Objective:***

To evaluate the PathoSEEK® *Pseudomonas aeruginosa* and *Staphylococcus aureus* Multiplex Detection Assay v2 using the MaGiC Lysis protocols with optional Grim Reeper for the presence/absence detection of *Pseudomonas aeruginosa* and *Staphylococcus aureus* species in cannabis flower (delta 9-tetrahydrocannabinol >0.3%; 1g) and marijuana-infused products (MIP).

### ***Results:***

Inclusivity and exclusivity results showed that the PathoSEEK® *Pseudomonas aeruginosa* and *Staphylococcus aureus* Multiplex Detection Assay v2 with MaGiC Lysis is highly specific in discriminating target organisms found in cannabis flower and infused products from non-target organisms. The kit was further verified by running NSI Certified Reference Materials, part numbers FM 628, 629, 632, and 635. Results show that the MaGiC Lysis kit and PathoSEEK® *Pseudomonas aeruginosa* and *Staphylococcus aureus* Multiplex Detection Assay v2 accurately detected *Pseudomonas aeruginosa* and *Staphylococcus aureus* when present.

## **Materials**

PathoSEEK® Pseudomonas aeruginosa and Staphylococcus aureus Multiplex Detection Assay v2 with MaGiC Lysis Kit

**P/N 420526** (Kit contains sufficient reagents for 200 reactions)

Kit Components:

Component Name	Qty Provided	Storage Conditions
MaGiC Lysis Reagent	1 Bottle (12 mL)	RT
MaGiC Stabilization Buffer	1 Bottle (24 mL)	RT
PathoSEEK® Amplification Mix	3 Vials (67 rxns/each)	RT / -20 °C*
PathoSEEK® Staph and Pseudo Multiplex Detection Assay v2	1 Tube (200 µL)	-20 °C

Additional Required Reagents Not in Kit:

Item P/N	Item Name	Qty Provided	Storage Conditions
420337	Internal Cannabis Control	1 Tube (50 µL)	-20 °C
420338	PathoSEEK® Staph and Pseudo Positive Control	1 Tube (50 µL)	-20 °C
420205	Tryptic Soy Broth	CS/10 x 500mL bottles	2-25°C
420184	PCR Grade Water	500 mL Bottle	2-25°C

## Inclusivity and Exclusivity

### Wet Laboratory Methodology

For the inclusivity evaluation, ATCC strains of *P. aeruginosa* and *S. aureus* were evaluated. Target strains were cultured in Tryptic Soy Broth for 16 hours at 37° C followed by MaGiC Lysis and qPCR with the PathoSEEK *P. aeruginosa* and *S. aureus* Multiplex Detection Assay v2.

For exclusivity, 37 organisms were evaluated. Target strains were cultured under optimal conditions for growth of the organism followed by MaGiC Lysis and qPCR with the PathoSEEK *P. aeruginosa* and *S. aureus* Multiplex Detection Assay v2.

### Results

Table 1: *Staphylococcus aureus* Inclusivity Results

Organism	ATCC Number	Origin	Result
<i>Staphylococcus aureus</i>	12600	Pleural fluid	x

Table 2: *Pseudomonas aeruginosa* Inclusivity Results

Organism	ATCC Number	Origin	Result
<i>Pseudomonas aeruginosa</i>	15442	Water bottle in animal room	x

Table 3: Exclusivity Results

Organism	ATCC Number	Origin	Results	
			Staphylococcus	Pseudomonas
<i>Rhizopus oryzae</i>	52748	N/A	ND	ND
<i>Pantoea agglomerans</i>	33243	N/A	ND	ND
<i>Penicillium chrysogenum</i>	10106	Cheese	ND	ND
<i>Aeromonas hydrophila</i>	7966	Tin of milk with fishy odor	ND	ND
<i>Proteus mirabilis</i>	43071	Rectum	ND	ND

<i>Citrobacter koseri</i>	25408	Throat	ND	ND
<i>Ralstonia insidiosa</i>	49129	Clinical isolate	ND	ND
<i>Salmonella enterica</i>	13311	Feces; food poisoning	ND	ND
<i>Pseudomonas fluorescens</i>	13525	Pre-filter tanks	ND	ND
<i>Purpureocillium lilacinum</i>	10114	Soil	ND	ND
<i>Bacillus subtilis</i>	23857	N/A	ND	ND
<i>Escherichia coli</i>	25922	Clinical isolate	ND	ND
<i>Aureobasidium pullulans</i>	62921	N/A	ND	ND
<i>Pseudomonas putida</i>	47054	N/A	ND	ND
<i>Escherichia hermannii</i>	700368	N/A	ND	ND
<i>Morganella morganii</i>	25829	Stool from Providence City Hospital	ND	ND
<i>Penicillium citrinum</i>	9849	N/A	ND	ND
<i>Klebsiella pneumoniae</i>	BAA-2146	Urine	ND	ND
<i>Trichosporon mucoides</i>	204094	N/A	ND	ND
<i>Burkholderia multivorans</i>	17616	Soil enriched with anthranilate at 41°C	ND	ND
<i>Scopulariopsis acremonium</i>	58636	Chicken house soil	ND	ND
<i>Erwinia rhapontici</i>	23376	Rheum rhaponticum	ND	ND
<i>Candida albicans</i>	10231	N/A	ND	ND
<i>Rahnella aquatilis</i>	33991	Man with bronchomycosis	ND	ND
<i>Meyerozyma guilliermondii</i>	6260	Sputum from patient with bronchomycosis	ND	ND
<i>Edwardsiella tarda</i>	23672	N/A	ND	ND
<i>Botrytis cinerea</i>	11542	Azalea flowers	ND	ND
<i>Citrobacter braakii</i>	BAA-3037	Urine	ND	ND
<i>Cryptococcus neoformans</i>	208821	Patient with Hodgkin's disease	ND	ND
<i>Hafnia alvei</i>	51873	Feces	ND	ND
<i>Papiliotrema laurentii</i>	18803	Palm wine, malaffou	ND	ND
<i>Klebsiella oxytoca</i>	51983	Blood	ND	ND
<i>Rhizopus stolonifer</i>	14037	N/A	ND	ND
<i>Aspergillus tamarii</i>	12669	Industrial pipe insulation	ND	ND
<i>Acinetobacter baumannii</i>	19606	Urine	ND	ND

<i>Aspergillus clavatus</i>	1007	N/A	ND	ND
<i>Aspergillus ustus</i>	1041	Culture contaminant	ND	ND

Note: Sanger sequencing of the assay amplicon confirmed cross-reactivity with *Pseudomonas rhizosphaerae*, an off-target species. This organism was not included in the original primer exclusion criteria, as it is not represented in common microbial reference collections such as ATCC. Because *P. rhizosphaerae* is not commercially available for purchase or propagation, it cannot be incorporated into formal exclusivity testing. To date, no clinical evidence has linked *P. rhizosphaerae* to human disease, although its presence in inhaled agricultural products warrants continued evaluation. Because *P. rhizosphaerae* is not commercially available from ATCC, its colony-forming characteristics cannot be readily assessed. As a result, the degree to which this species may contribute to inflated CFU/g estimates in the *P. aeruginosa* assay remains undetermined.

### **Limit of Detection**

Cultures of *S. aureus* and *P. aeruginosa* were grown for 16 hours. After 16 hours a series of six 1:10 dilutions were made. Per the MaGiC Lysis kit's instructions, 10 µL of each dilution in triplicate was lysed using the MaGiC lysis buffer. The dilutions were also plated on 3M Rapid Aerobic Count (RAC) plates for comparison. For plating on RAC petrifilm, 10 µl of the appropriate dilution was added to 990ul of water, keeping it consistent with the amount of enrichment going into the MaGiC Lysis, and 1 mL was added to the RAC plates and then incubated at 37 °C for 24 hours. The results can be found below in Table 4.

Table 4: Limit of Detection and Plate Count Comparison Results

Sample	Dilution	MaGiC Lysis				3M RAC Plate
		Target Fam	Cq Fam	Target Rox	Cq Rox	CFU Count
<i>Pseudomonas</i> Culture	1:10	<i>Staphylococcus</i>	ND	<i>Pseudomonas</i>	22.52	3,220,000
<i>Pseudomonas</i> Culture	1:100	<i>Staphylococcus</i>	ND	<i>Pseudomonas</i>	26.90	322,000
<i>Pseudomonas</i> Culture	1:1,000	<i>Staphylococcus</i>	ND	<i>Pseudomonas</i>	30.08	32,200
<i>Pseudomonas</i> Culture	1:10,000	<i>Staphylococcus</i>	ND	<i>Pseudomonas</i>	32.97	3,220
<i>Pseudomonas</i> Culture	1:100,000	<i>Staphylococcus</i>	ND	<i>Pseudomonas</i>	36.31	322
<i>Pseudomonas</i> Culture	1:1,000,000	<i>Staphylococcus</i>	ND	<i>Pseudomonas</i>	38.39	30

<i>Staphylococcus</i> Culture	1:10	<i>Staphylococcus</i>	24.26	<i>Pseudomonas</i>	ND	<i>1,460,000</i>
<i>Staphylococcus</i> Culture	1:100	<i>Staphylococcus</i>	27.91	<i>Pseudomonas</i>	ND	<i>146,000</i>
<i>Staphylococcus</i> Culture	1:1,000	<i>Staphylococcus</i>	31.26	<i>Pseudomonas</i>	ND	<i>14,600</i>
<i>Staphylococcus</i> Culture	1:10,000	<i>Staphylococcus</i>	34.90	<i>Pseudomonas</i>	ND	<i>1,460</i>
<i>Staphylococcus</i> Culture	1:100,000	<i>Staphylococcus</i>	38.17	<i>Pseudomonas</i>	ND	146
<i>Staphylococcus</i> Culture	1:1,000,000	<i>Staphylococcus</i>	38.77	<i>Pseudomonas</i>	ND	16

The italicized numbers were too numerous to count and extrapolated from the higher dilutions.

### **Proficiency Testing / Certified Reference Material**

Four different CRMs were tested:

(*S. aureus* in hemp and oil, and *P. aeruginosa* in hemp and edible). 9 mL of TSB was added to each vial then incubated at 37 °C for 24 hours. After enrichment, the MaGiC Lysis protocol was followed and the results can be found below in Table 5.

Table 5: NSI CRM Data

CRM	Catalogue #	Lot #	Dilution	Target Fam	Cq FAM	Target Hex	Cq HEX	Target Cy5	Cq Cy5
<i>S. aureus</i> Hemp	FM-628	240913	Straight	<i>S. aureus</i>	+	ICC	+	<i>P. aeruginosa</i>	ND
<i>S. aureus</i> Hemp	FM-628	240913	Straight	<i>S. aureus</i>	+	ICC	+	<i>P. aeruginosa</i>	ND
<i>S. aureus</i> Hemp	FM-628	240913	Straight	<i>S. aureus</i>	+	ICC	+	<i>P. aeruginosa</i>	ND
<i>S. aureus</i> Oil	FM-632	240307	Straight	<i>S. aureus</i>	+	ICC	+	<i>P. aeruginosa</i>	ND
<i>S. aureus</i> Oil	FM-632	240307	Straight	<i>S. aureus</i>	+	ICC	+	<i>P. aeruginosa</i>	ND
<i>S. aureus</i> Oil	FM-632	240307	Straight	<i>S. aureus</i>	+	ICC	+	<i>P. aeruginosa</i>	ND
<i>P. aeruginosa</i> Hemp	FM-629	230713	1:10	<i>S. aureus</i>	ND	ICC	+	<i>P. aeruginosa</i>	+
<i>P. aeruginosa</i> Hemp	FM-629	230713	1:10	<i>S. aureus</i>	ND	ICC	+	<i>P. aeruginosa</i>	+
<i>P. aeruginosa</i> Hemp	FM-629	230713	1:10	<i>S. aureus</i>	ND	ICC	+	<i>P. aeruginosa</i>	+
<i>P. aeruginosa</i> Edible	FM-635	230831	1:10	<i>S. aureus</i>	ND	ICC	+	<i>P. aeruginosa</i>	+
<i>P. aeruginosa</i> Edible	FM-635	230831	1:10	<i>S. aureus</i>	ND	ICC	+	<i>P. aeruginosa</i>	+
<i>P. aeruginosa</i> Edible	FM-635	230831	1:10	<i>S. aureus</i>	ND	ICC	+	<i>P. aeruginosa</i>	+
Positive Control				<i>S. Aureus</i>	14.85	ICC	ND	<i>P. aeruginosa</i>	16.55
NTC				<i>S. Aureus</i>	ND	ICC	ND	<i>P. aeruginosa</i>	ND

### **Grim Reefer Free DNA Removal Validation**

To ensure that Grim Reefer free DNA removal works with PathoSEEK *P. aeruginosa* and *S. aureus* Multiplex Assay v2, live organisms were enriched in the presence of flower, oil, and chocolate for 16 hours. After 16 hours samples were tested with and without the optional Grim Reefer step according to the MaGiC Lysis protocol. The target organisms were detected in the inoculated matrices with and without Grim Reefer, which can be seen in Table 6.

Table 6: Grim Reefer Comparison Data

Sample	Matrix	Grim Reefer	Target Fam	Cq Fam	Target Hex	Cq Hex	Target Cy5	Cq Cy5
<i>Pseudomonas aeruginosa</i>	Flower	No	<i>S. aureus</i>	ND	IC	+	<i>P. aeruginosa</i>	+
<i>Pseudomonas aeruginosa</i>	Flower	Yes	<i>S. aureus</i>	ND	IC	+	<i>P. aeruginosa</i>	+
<i>Staphylococcus aureus</i>	Flower	No	<i>S. aureus</i>	+	IC	+	<i>P. aeruginosa</i>	ND
<i>Staphylococcus aureus</i>	Flower	Yes	<i>S. aureus</i>	+	IC	+	<i>P. aeruginosa</i>	ND
<i>Pseudomonas aeruginosa</i>	Chocolate	No	<i>S. aureus</i>	ND	IC	+	<i>P. aeruginosa</i>	+
<i>Pseudomonas aeruginosa</i>	Chocolate	Yes	<i>S. aureus</i>	ND	IC	+	<i>P. aeruginosa</i>	+
<i>Staphylococcus aureus</i>	Chocolate	No	<i>S. aureus</i>	+	IC	+	<i>P. aeruginosa</i>	ND
<i>Staphylococcus aureus</i>	Chocolate	Yes	<i>S. aureus</i>	+	IC	+	<i>P. aeruginosa</i>	ND
<i>Pseudomonas aeruginosa</i>	Oil	No	<i>S. aureus</i>	ND	IC	+	<i>P. aeruginosa</i>	+
<i>Pseudomonas aeruginosa</i>	Oil	Yes	<i>S. aureus</i>	ND	IC	+	<i>P. aeruginosa</i>	+
<i>Staphylococcus aureus</i>	Oil	No	<i>S. aureus</i>	+	IC	+	<i>P. aeruginosa</i>	ND
<i>Staphylococcus aureus</i>	Oil	Yes	<i>S. aureus</i>	+	IC	+	<i>P. aeruginosa</i>	ND
Positive Control			<i>S. aureus</i>	14.32	IC	ND	<i>P. aeruginosa</i>	16.41
NTC			<i>S. aureus</i>	ND	IC	ND	<i>P. aeruginosa</i>	ND

### **Conclusions**

The MaGiC Lysis kit in conjunction with the PathoSEEK® *Pseudomonas aeruginosa* and *Staphylococcus aureus* Multiplex Detection Assay v2 is a rapid, alternative method to traditional plating procedures for accurate detection of *Pseudomonas Aeruginosa* and *Staphylococcus aureus* in cannabis flower and cannabis infused products.

## REVISION HISTORY

Version	Date	Description
v1	December 2024	Validation for MaGiC Lysis with <i>Pseudomonas aeruginosa</i> and <i>Staphylococcus aureus</i> Multiplex Assay v2 Method
v2	June 2025	Addition of validation data for Grim Reefer Free DNA Removal & note about exclusivity data concerning <i>P. rhizosphaerae</i>

## DISCLAIMER

This test was developed, and its performance characteristics determined by Medicinal Genomics Company, for laboratory use. Any deviations from this protocol are not supported by MGC.

This test has not been validated on remediated (irradiated, ozone treated, acid treated, hydrogen peroxide treated, etc.) samples. Samples that have undergone remediation may cause discordant results between plating methods and PathoSEEK methods. When remediated samples produce a result above the action limit on qPCR, we recommend confirming viability with an approved plating method.

Results may vary based on laboratory conditions. Altitude and humidity are factors known to affect the growth of bacterial and fungal species.

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